

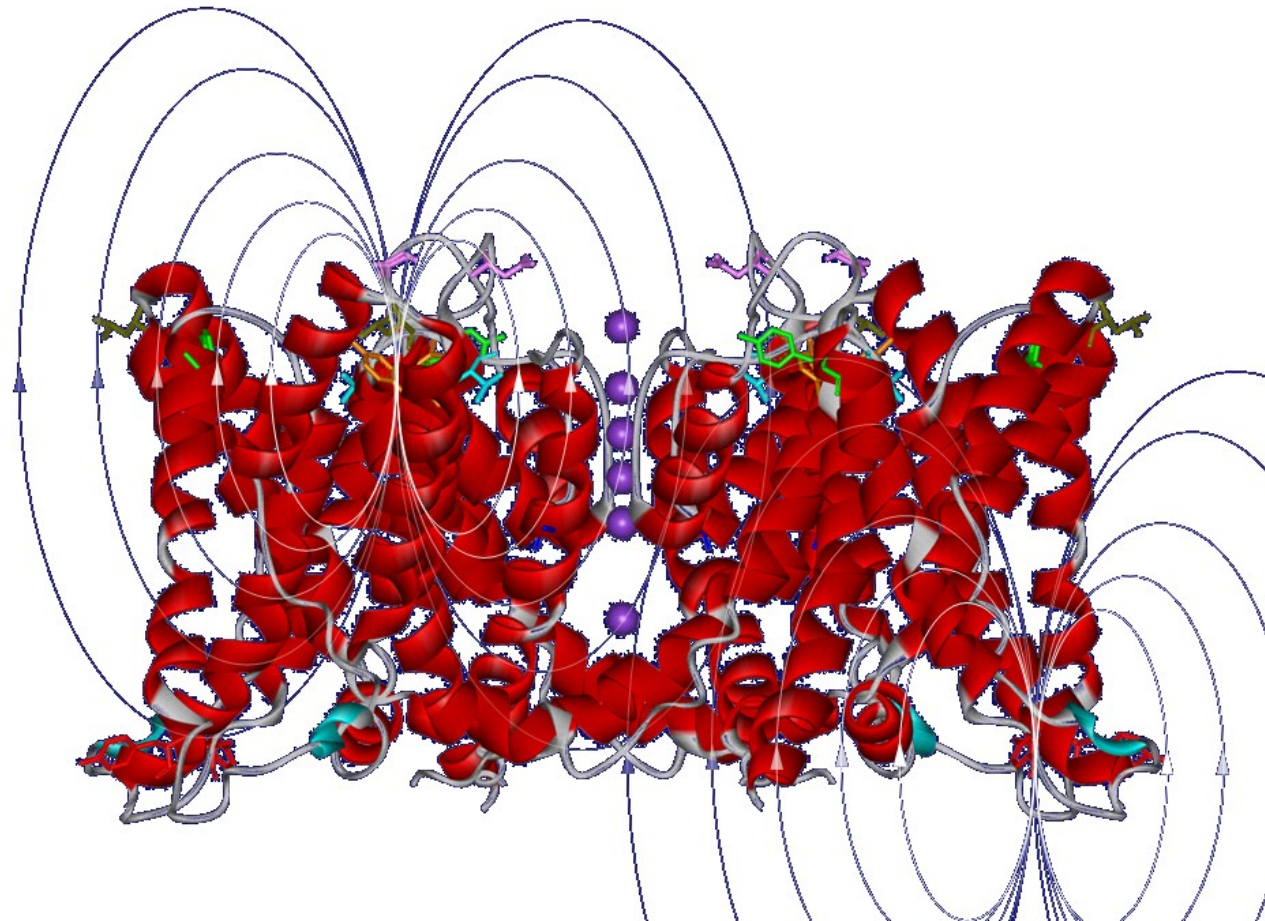
# Pulsed Electron Electron Double Resonance (PELDOR oder DEER)

**T. F. Prisner**

Institute of Phys. & Theor. Chemistry

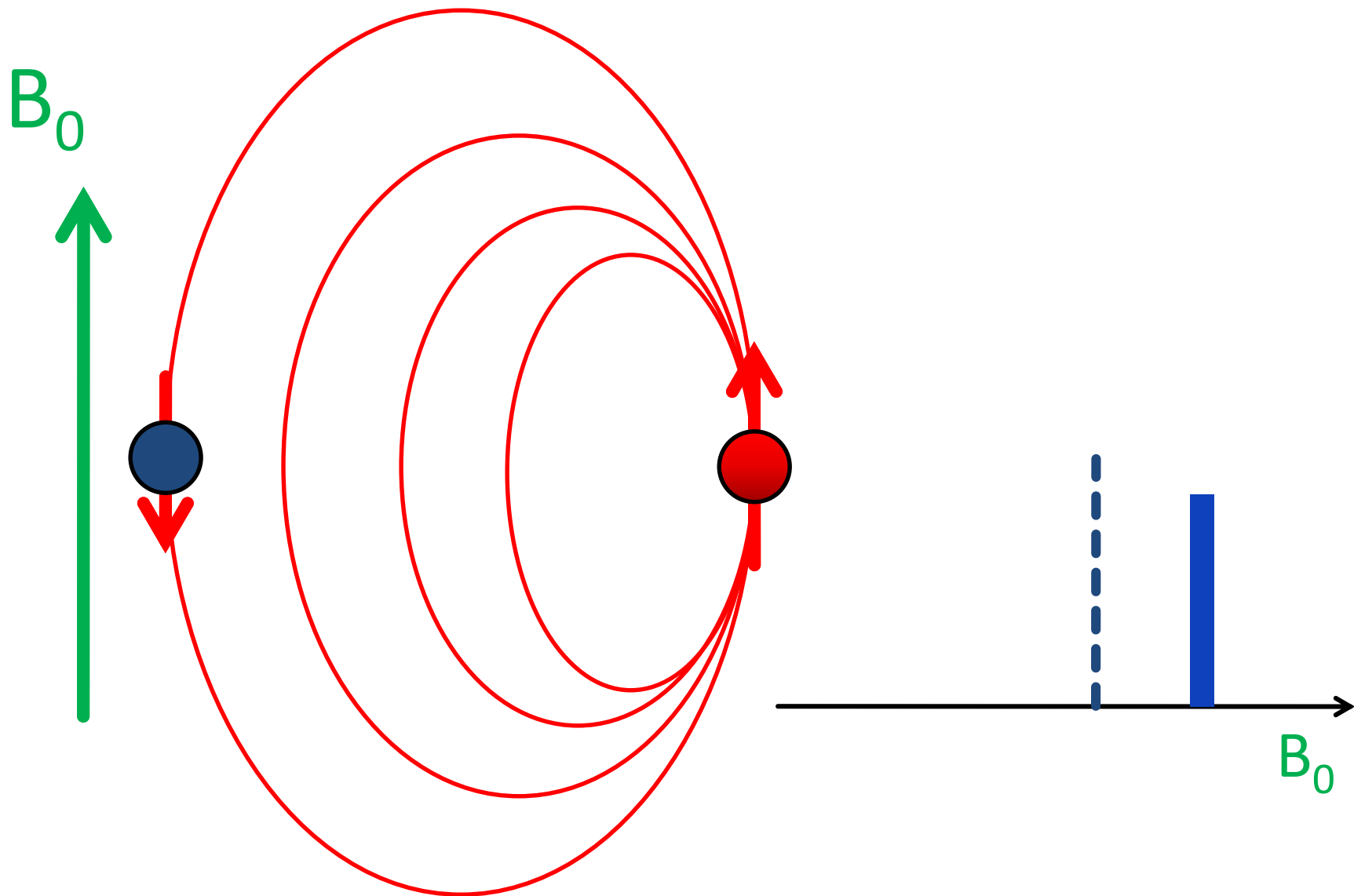
Center of Biological Magnetic Resonance

Goethe University Frankfurt



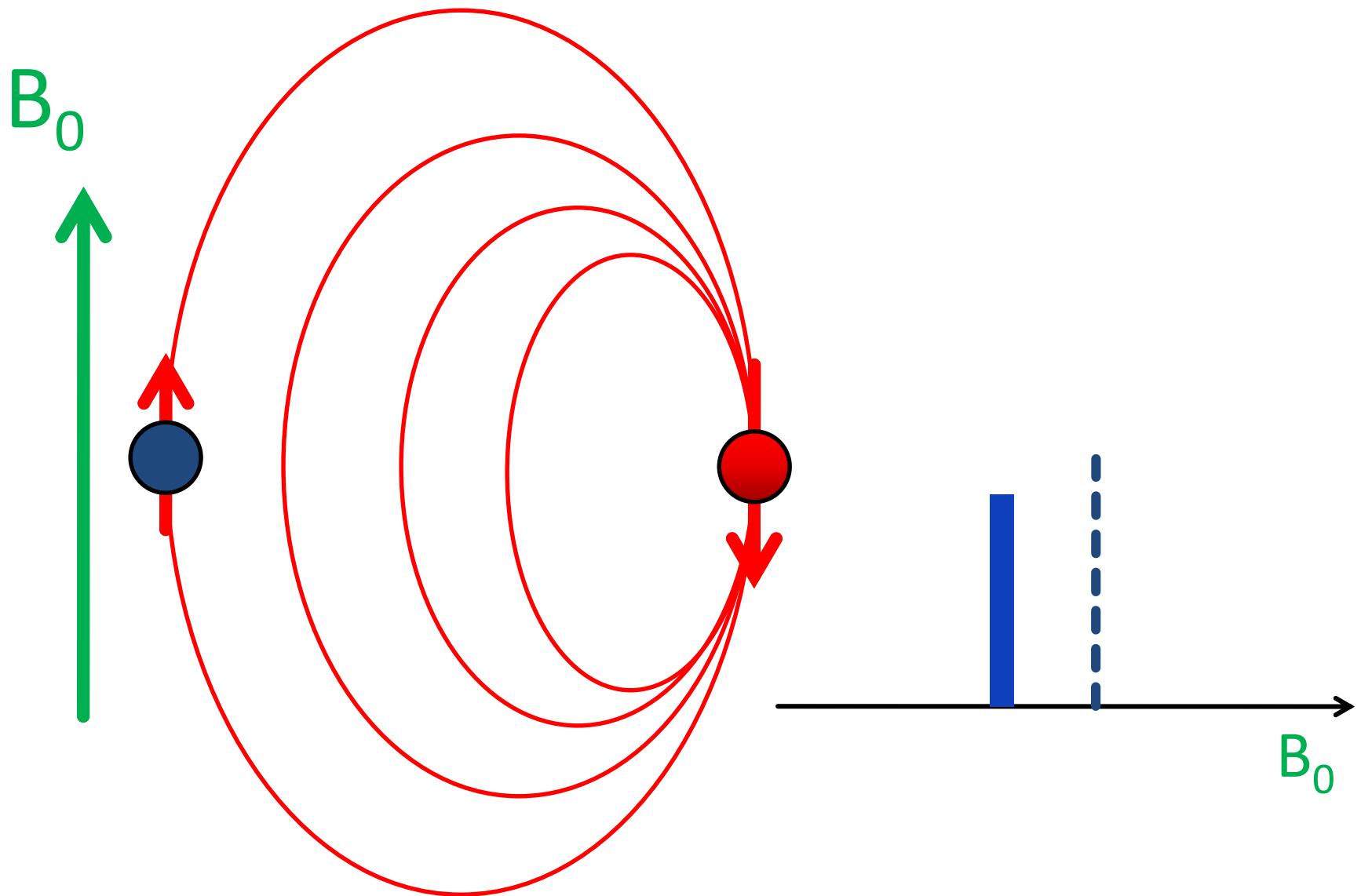
# Magnetic dipole-dipole interaction between two spin labels

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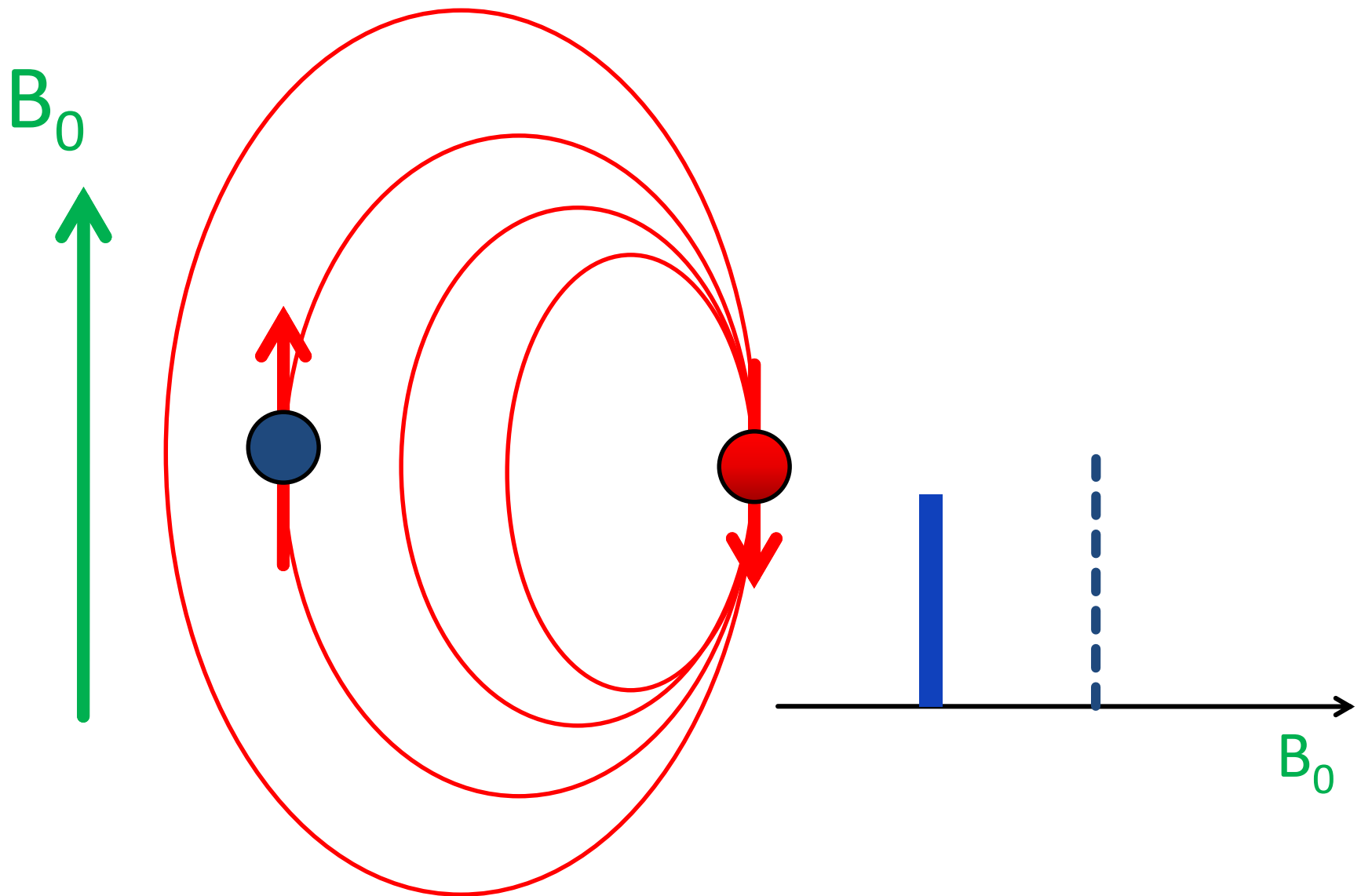
# Magnetic dipole-dipole interaction between two spin labels

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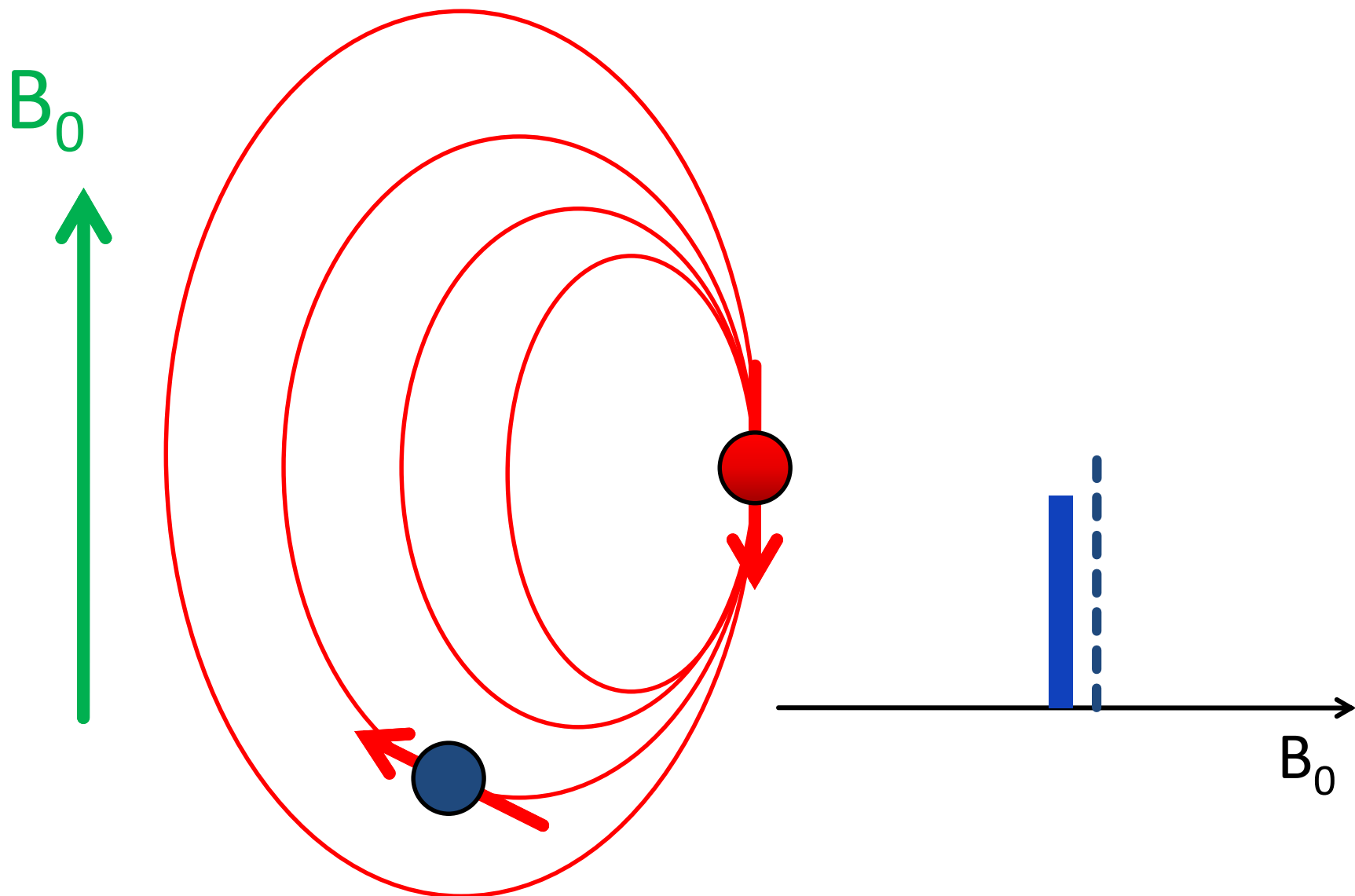
# Magnetic dipole-dipole interaction between two spin labels

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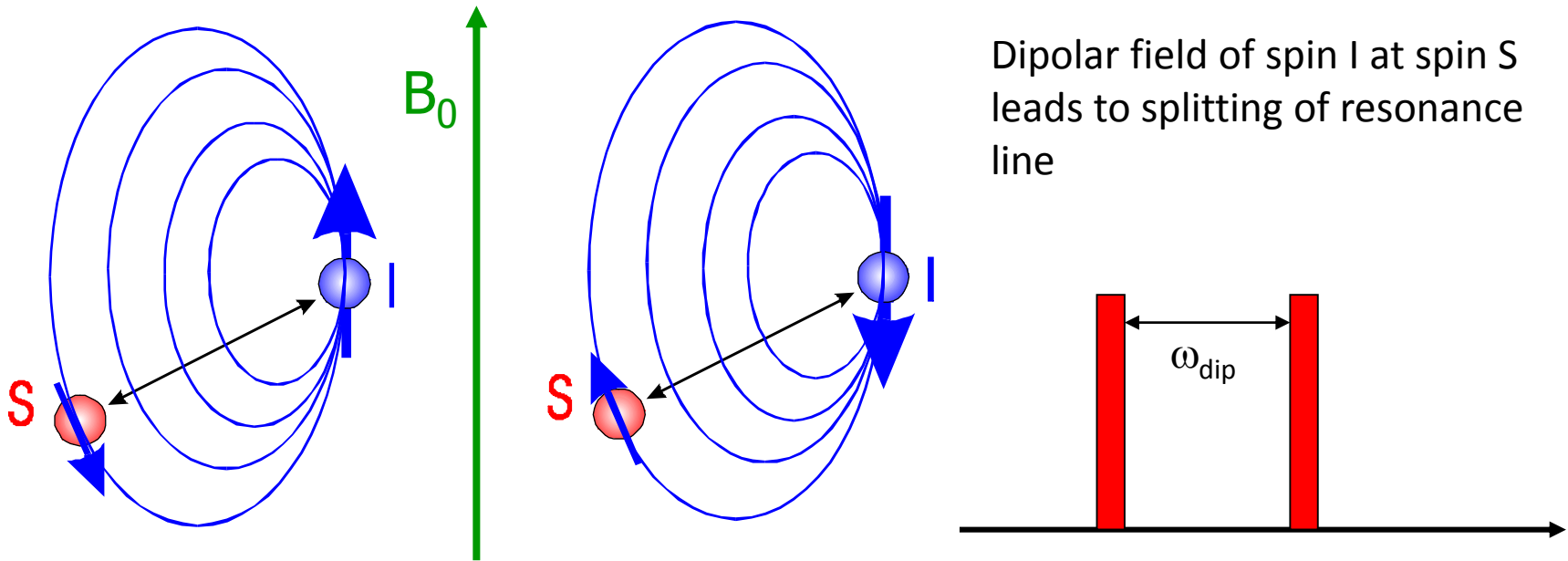


# Magnetic dipole-dipole interaction between two spin labels

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# Magnetic Dipol-Dipol Interaction leads to splitting



$$\omega_{dip} = \frac{\gamma_s \mu_I}{r_{SI}^3} (1 - 3 \cos^2(\theta)) \propto \frac{g_s^{eff} g_I^{eff}}{r_{SI}^3} (1 - 3 \cos^2(\theta))$$

Dipolar splitting depends on:

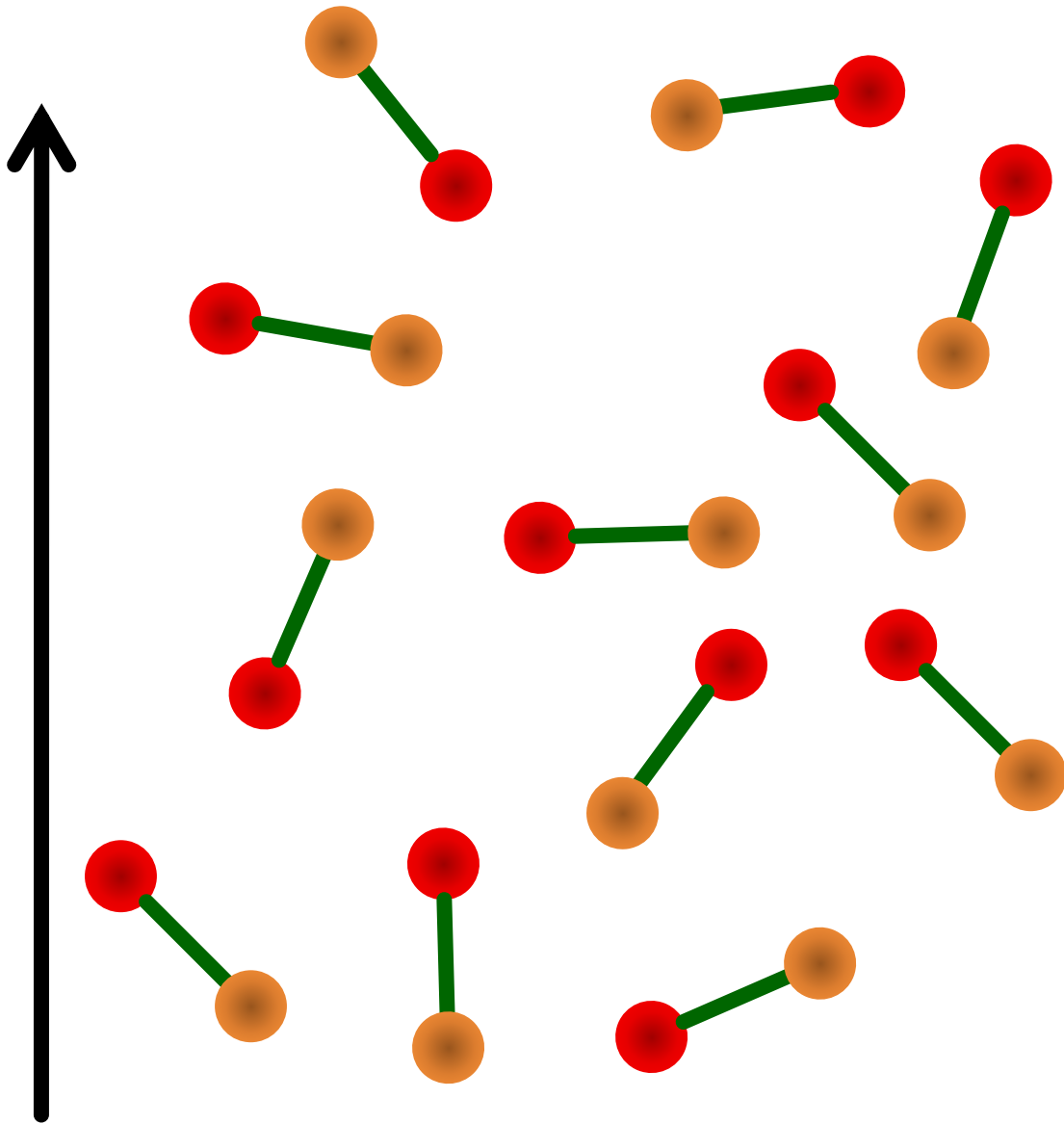
g values

orientation  $\theta$

distance R

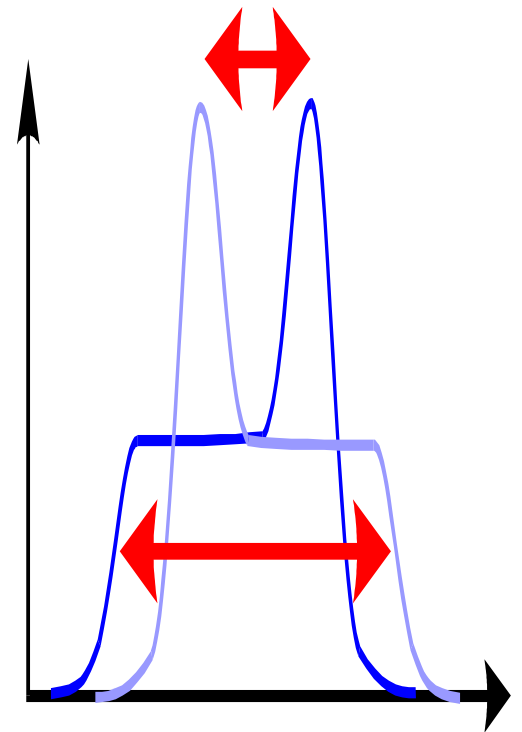
# Frozen Sample with random orientation of molecules

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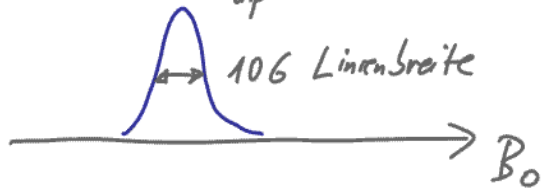
Dipolar Pake  
Pattern

Width < MHz

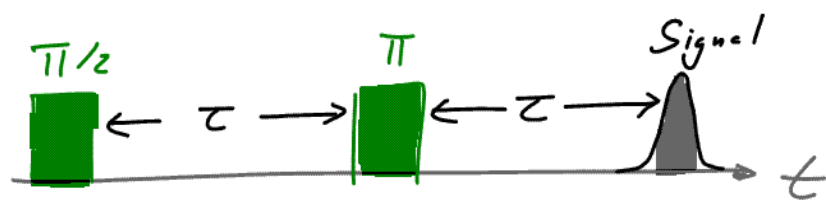


Bei  $R = 3 \text{ nm}$

$\leftrightarrow \Delta B_{\text{dip}} < 1 \text{ G}$  nicht detektierbar!

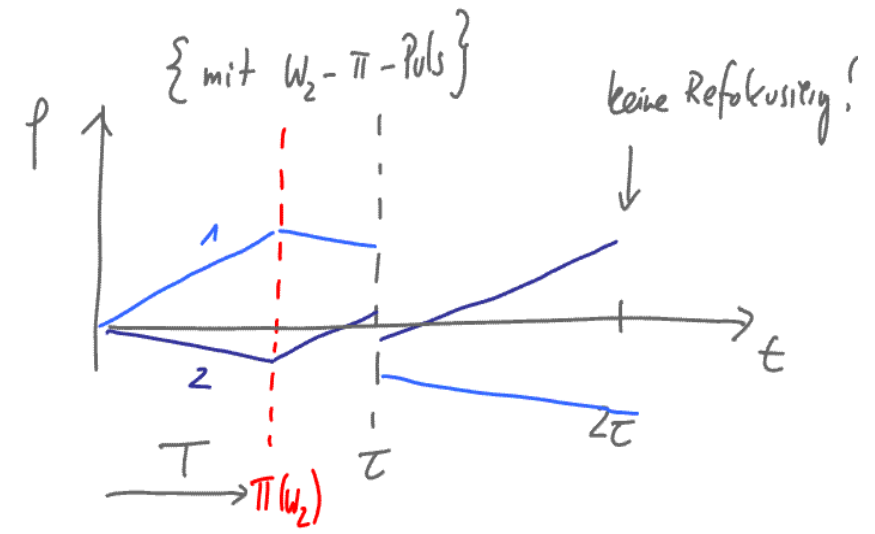
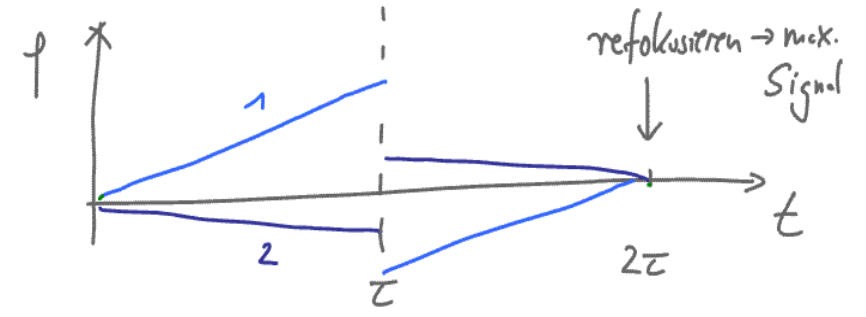
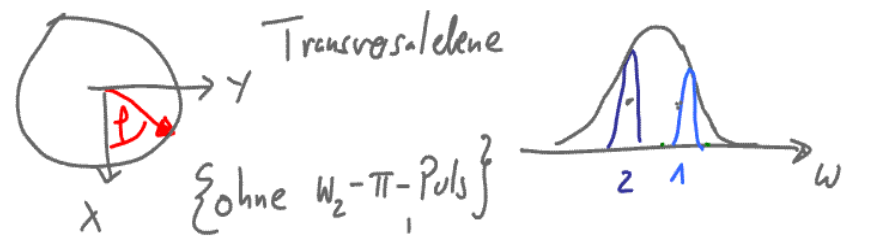


Idee 1: Benutze Pulsmethoden um inhomogene Linienbreite zu vermindern!



Problem: Refokussiert alle inhom. WW - also auch  $2\omega_{dd}$ !

Idee 2: Invertiere selektiv Spin 2 während dem Experiment  $\rightarrow$  verhindert Refokussierung von dipolarer WW

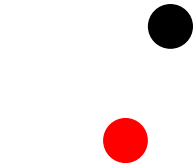
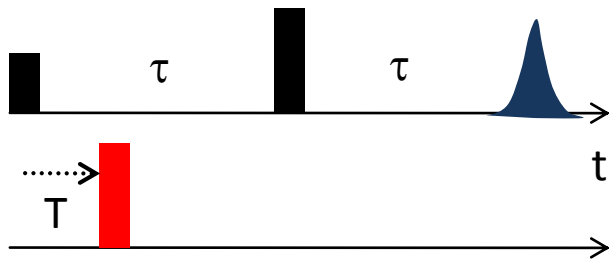


$$S(T) = S_{\text{max}} \cdot \cos(\omega_{dd} T)$$



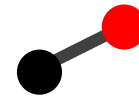
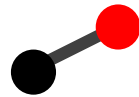
# Pulsed Electron Electron Double Resonance (PELDOR)

## 3-Pulse PELDOR Sequence



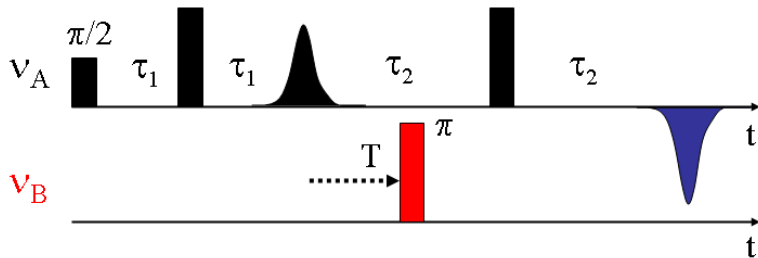
Milov, Salikov, Shirov, *Fiz.Tverd. Tela* 23, 975 (1981)  
Milov, Ponomarev, Tsvetkov, *Chem. Phys. Lett.* 110, 67 (1984)

## DEER (Double Electron-Electron Resonance)



Larsen, Singel, *J. Chem. Phys.* 98, 5134 460 (1993)

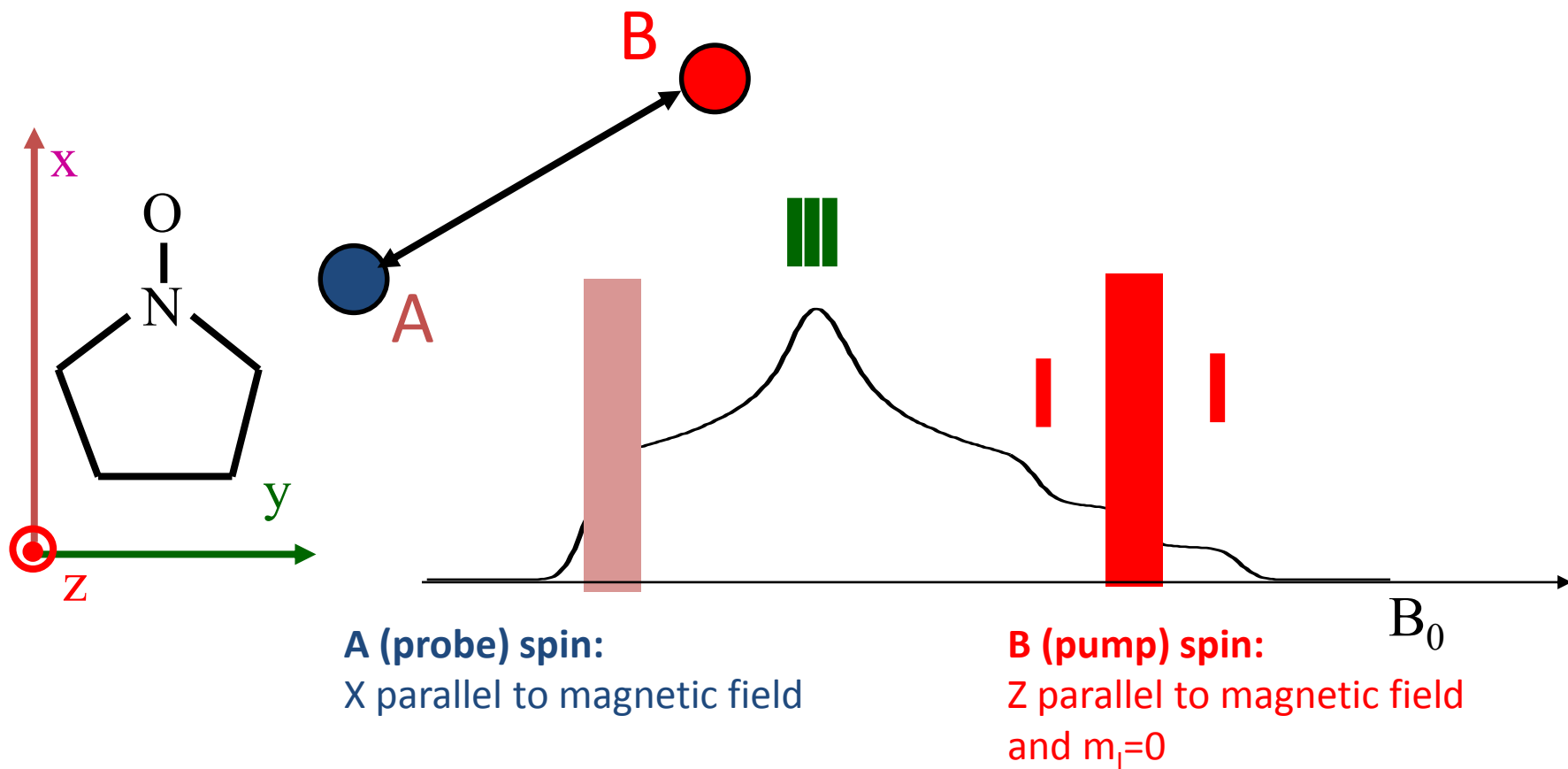
## 4-Pulse DEER Sequence



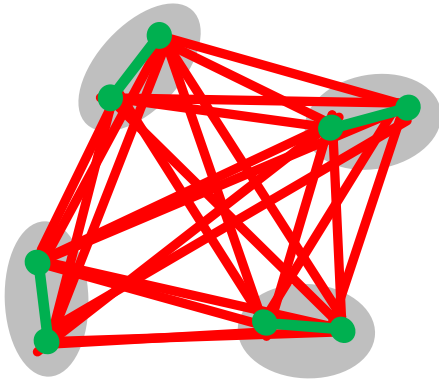
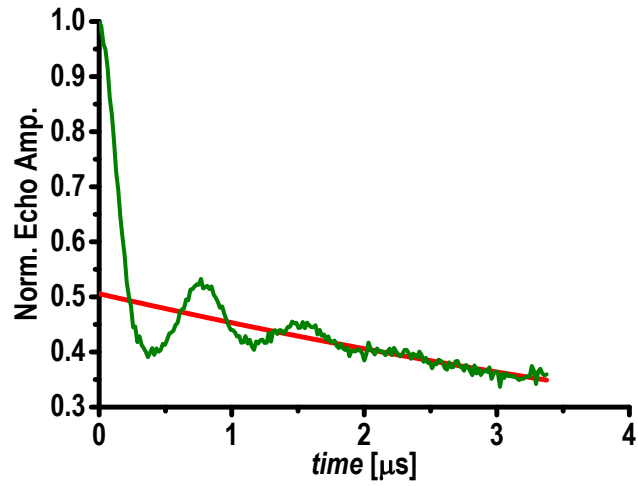
Martin, Pannier, Diederich, Gramlich, Hubrich, Spiess, *Angew. Chem.* 37, 2833 (1998)

Pannier, Veit, Godt, Jeschke, Spiess, *J. Magn. Reson.* 142, 331 (2000).

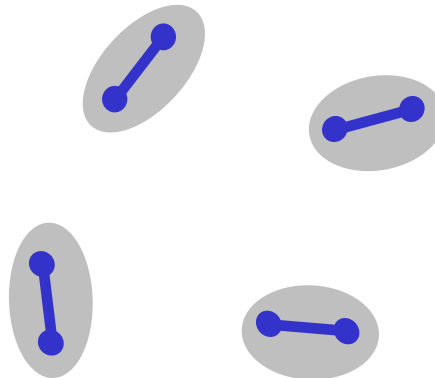
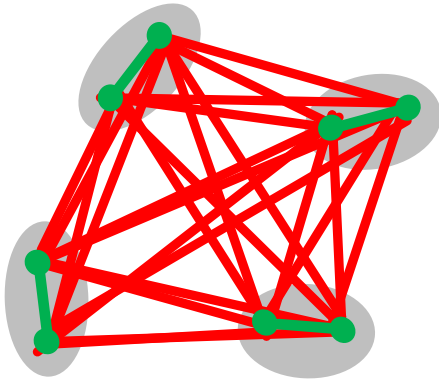
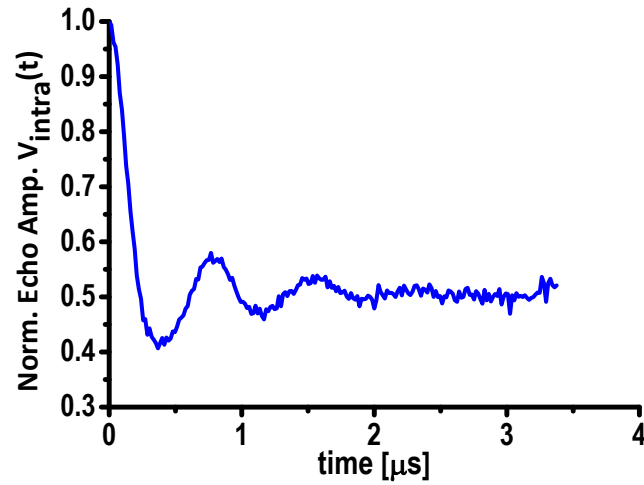
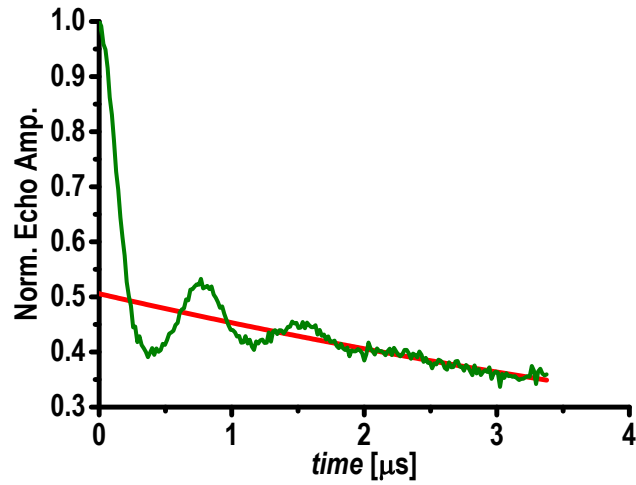
# How to define A and B spins ?



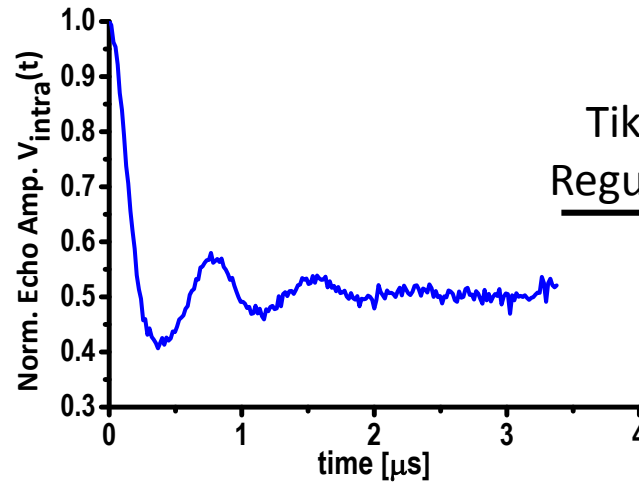
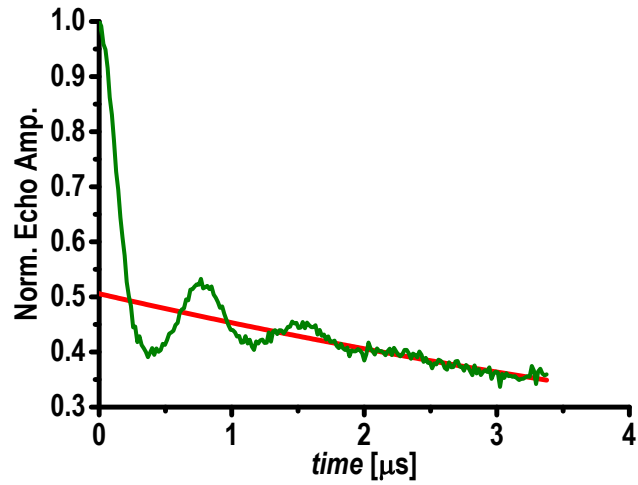
# PELDOR – Data Analysis



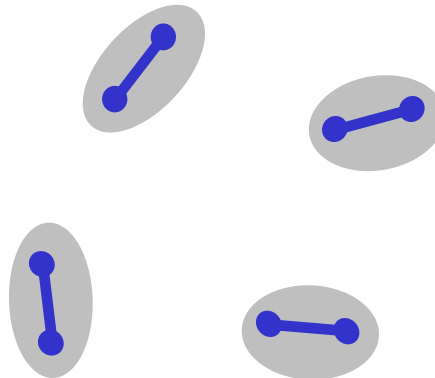
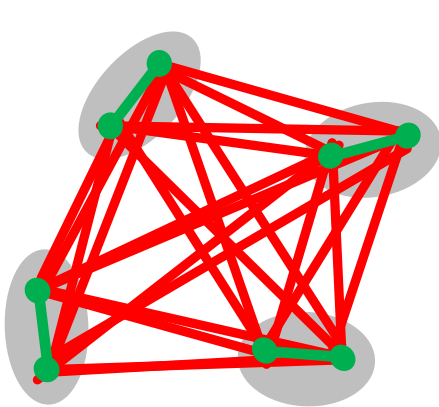
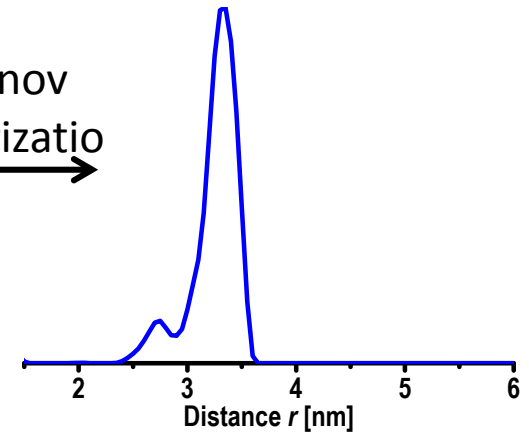
# PELDOR – Data Analysis



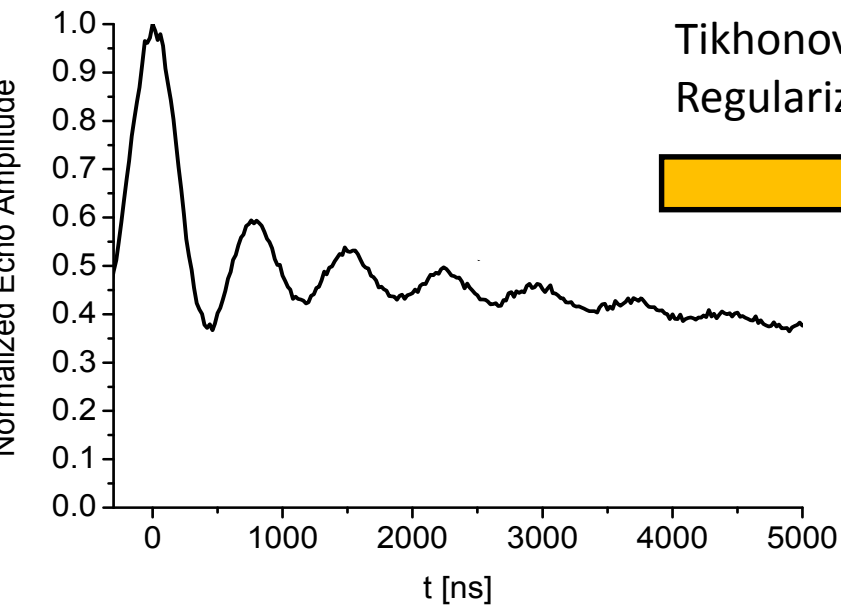
# PELDOR – Data Analysis



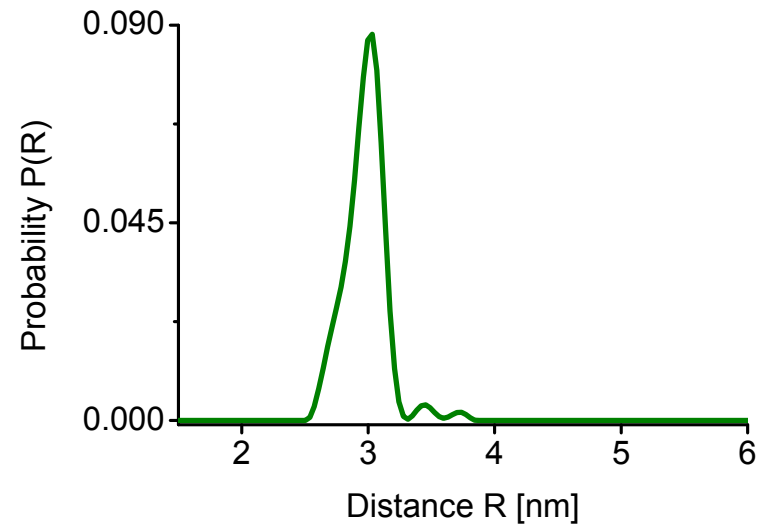
Tikhonov  
Regularizatio  
n



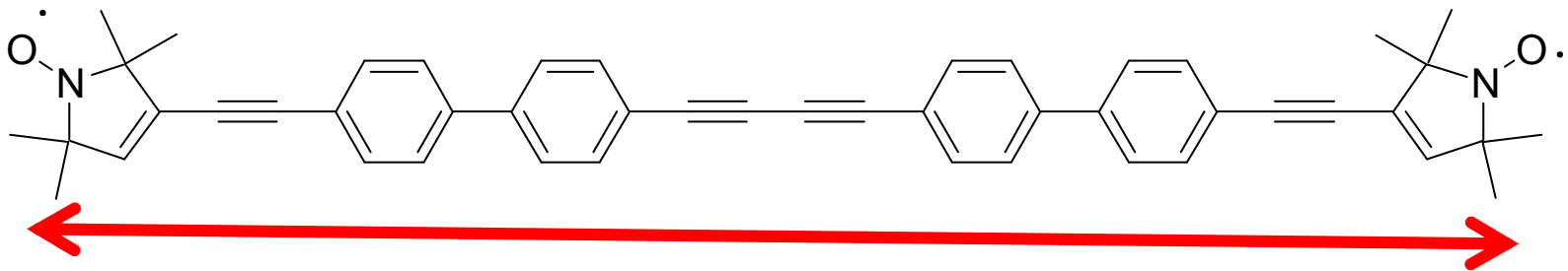
# Extraction of Distance Distribution Function



Tikhonov  
Regularization



- Fit of Pake-Patterns to Time trace
- Assumes  $R \geq 0$   $P(R) \geq 0$ , smooth function

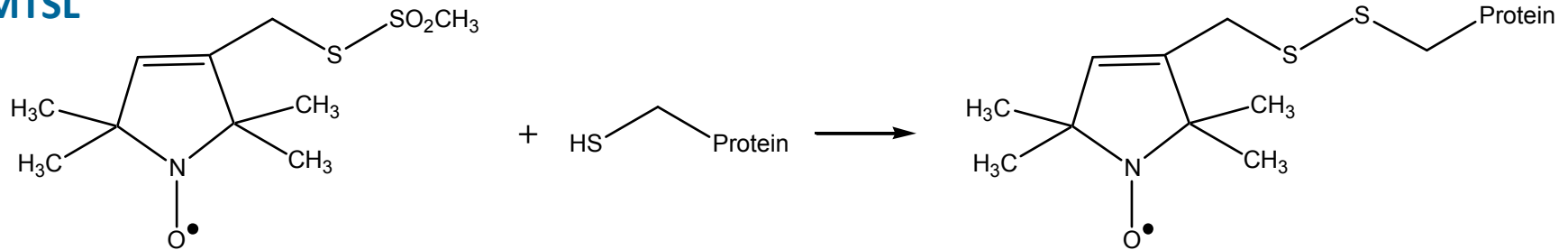


# Spin Labeling of Proteins

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Methanthiosulfonate:

**MTSL**



Auch:

Gd-Komplexe als Spin Labels (Säure-Resistent für in-cell Anwendungen)

Tryl-Radikale als Spin Label (eventuell für Experimente bei RT)

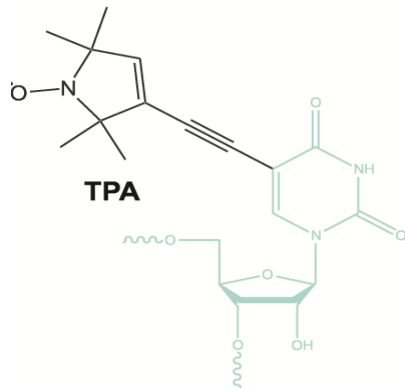
Oder:

Intrinsische Kofaktoren in Proteinen die sich reduzieren/oxidieren lassen oder per se paramagnetisch sind

(Ni, Cu, Mn, FeS, heme, aminosäure radikale, Flavine ....)

# Spin labels for Nucleic Acids

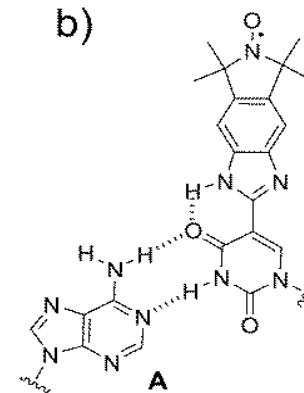
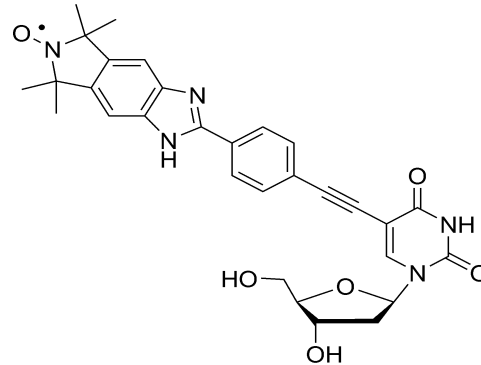
Joachim Engels  
(Uni Frankfurt)



Rotate around triple bond

*Nature Protocoll* (2007)  
*NAR* (2007)

Snorri Sigurdsson (University of Iceland)



Rigid in double stranded structures

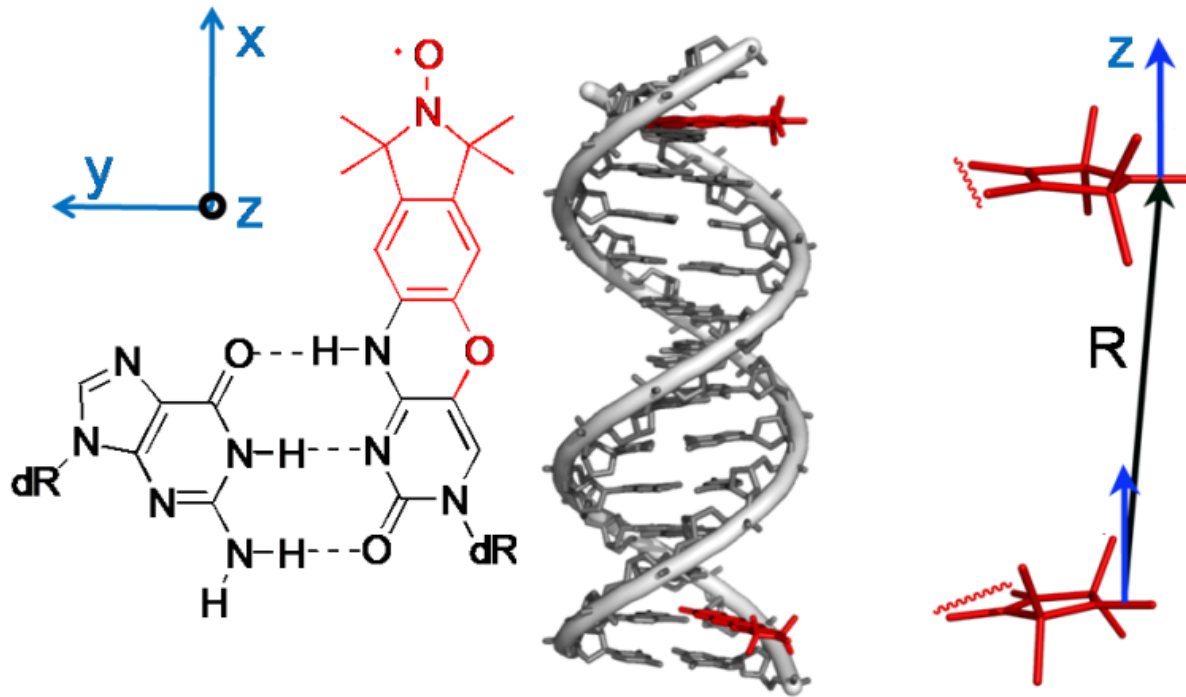
*Chem. Eur. J.* (2014)

*Angewandte* (2009)  
*JACS* (2011)

- All nucleotides in DNA and RNA can be spin-labeled
- Different degrees of rotational flexibility of spin label
- Different orientation with respect to nucleotide
- Synthesis with modified nucleobase



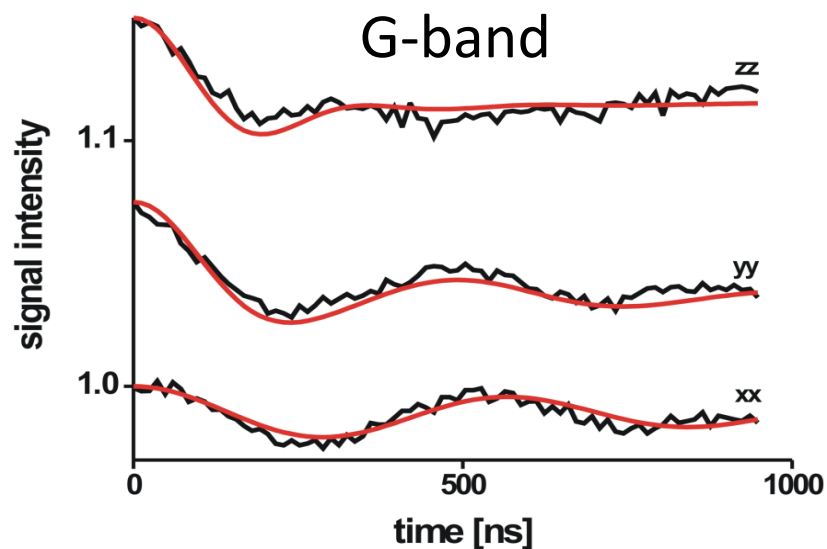
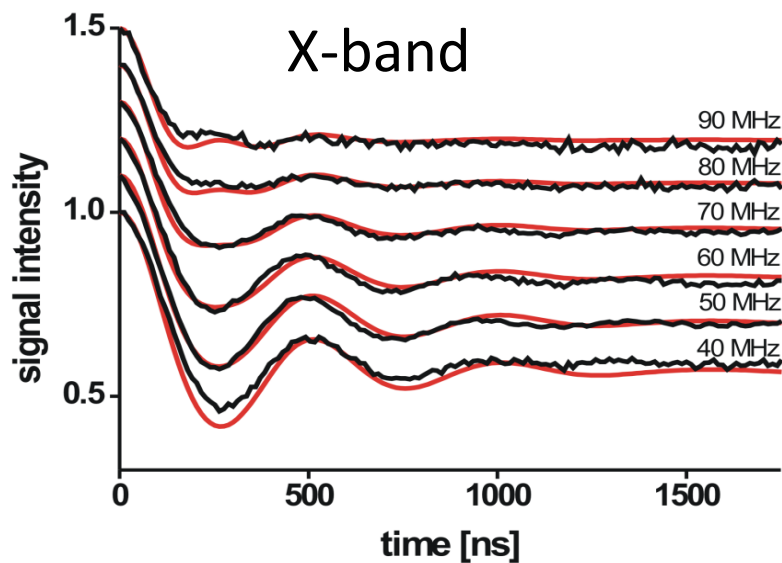
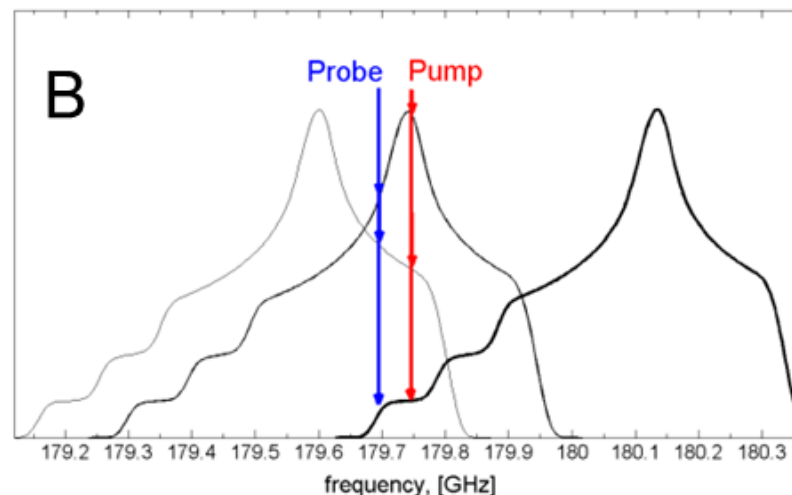
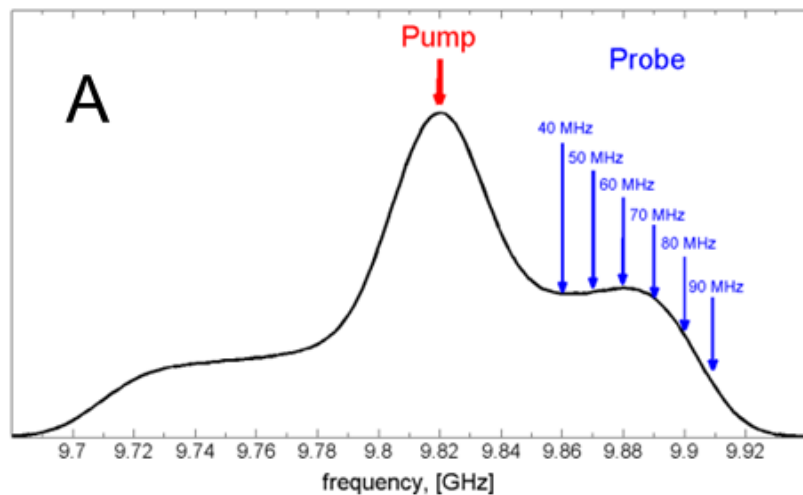
# Incorporation of rigid spin label into NAs



**Snorri Sigurdsson**  
(Reykjavic)

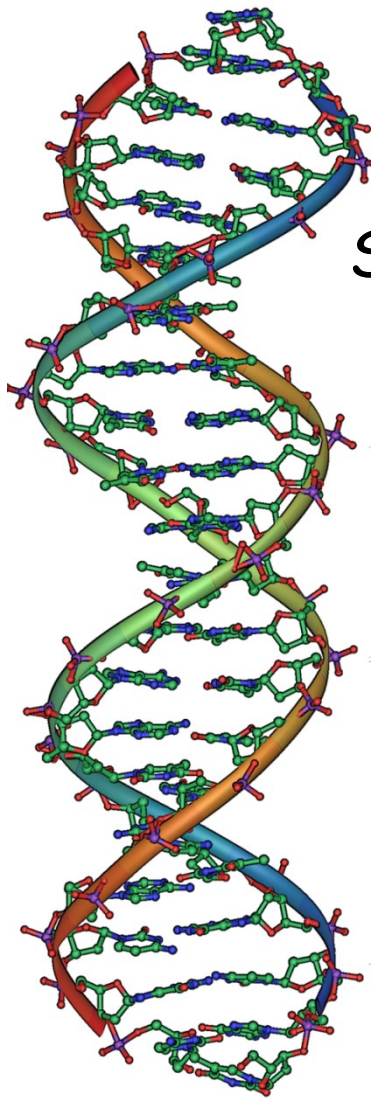
- Rigid in double stranded DNA and RNA
- Determination of orientation between two spin labels

# Multi-Frequency / Multi-Field PELDOR



# Investigation of dynamics of dsDNA

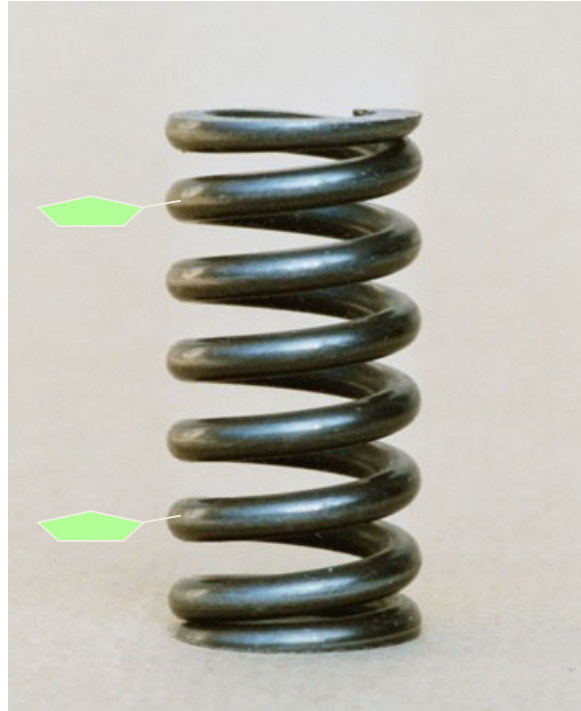
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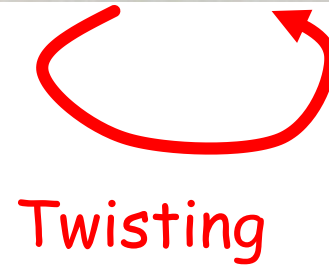
Stretching



$$\Delta x^2 \sim L$$

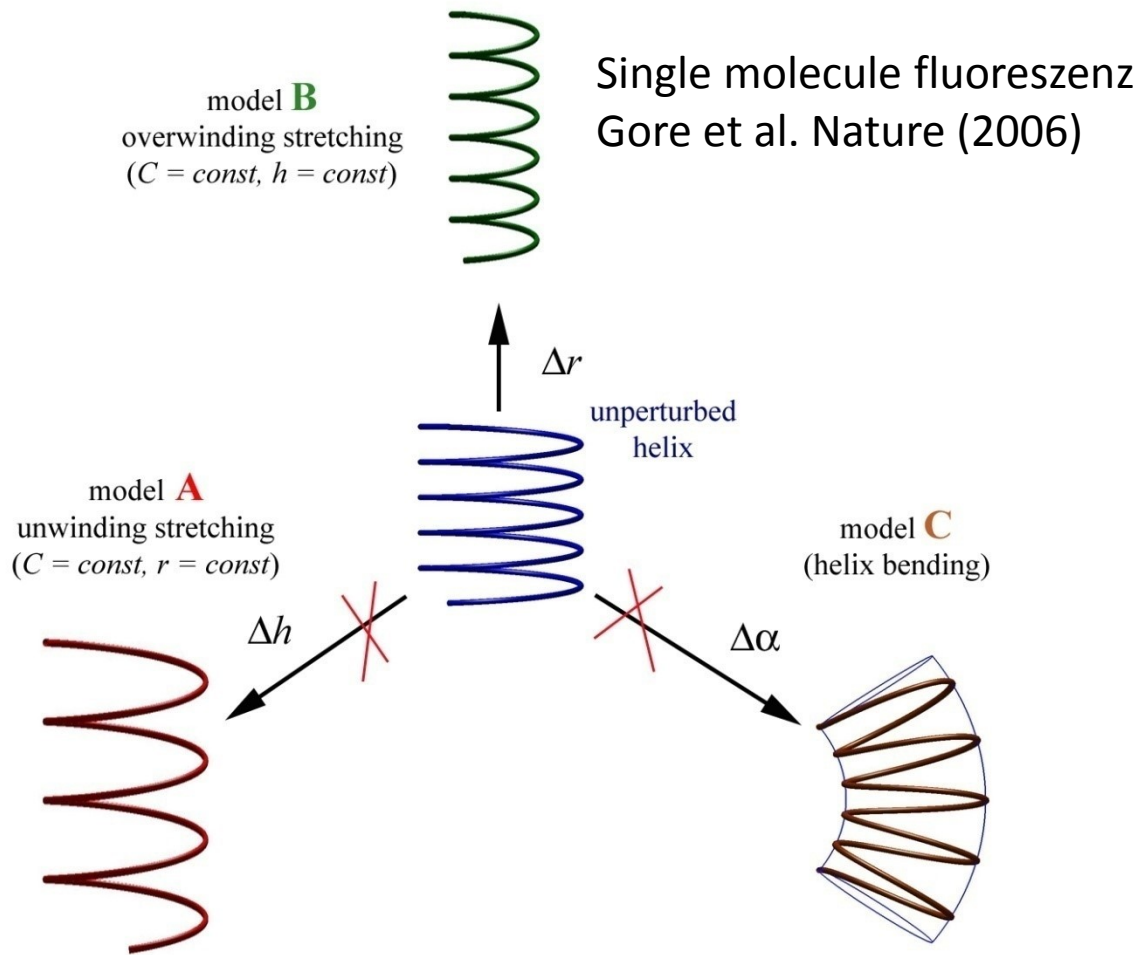


Bending

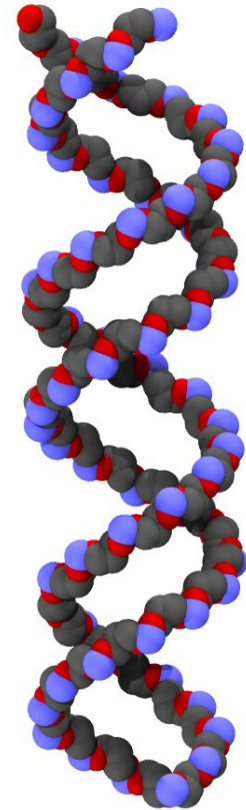


Twisting

# Models for conformational dynamics of dsDNA



Single molecule fluorescence  
Gore et al. Nature (2006)



PCA Grubmüller  
Bouvier & Grubmüller  
Biophys. J 93, 770 (2007)

SAXSI  
Mathew-Fenn et al. Science (2008)

Modeling  
Becker et al. Science (2009)

## **Übersichtsartikel zu EPR:**

Vorlesungsskript Gunnar Jeschke (ETH Zürich / MPI Mainz)

Jeschke/Schweiger Principles of Pulse EPR Oxford University Press

Lund/Shiotani/Shimada Principles and Applications of EPR Spectroscopy Springer

## **Übersichtsartikel zu PELDOR /DEER:**

Schiemann, O. and Prisner, T.F. (2007) Long-range distance determinations in biomacromolecules by EPR spectroscopy. *Quarterly Review in Biophysics*, **40**, 1-53.

Jeschke, G. (2012) *Annual Review of Physical Chemistry*, **63**, 419-446.

## **Übersichtsartikel zu PELDOR an RNA/DNA:**

EPR Spectroscopy (Topics in Current Chemistry 321, 2012)  
Springer Verlag

RNA Structure and Folding (Biophysical Techniques and Prediction Methods) 2013  
De Gruyter Verlag